

# Package ‘BiocSklern’

October 16, 2019

**Title** interface to python sklern via Rstudio reticulate

**Description** This package provides interfaces to selected sklern elements, and demonstrates fault tolerant use of python modules requiring extensive iteration.

**Version** 1.6.0

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**Suggests** testthat, restfulSE, HDF5Array, BiocStyle

**Depends** R (>= 3.5.0), reticulate, methods, SummarizedExperiment, knitr

**Imports** BBmisc

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**License** Artistic-2.0

**LazyLoad** yes

**biocViews** StatisticalMethod, DimensionReduction, Infrastructure

**RoxygenNote** 6.1.1

**VignetteBuilder** knitr

**SystemRequirements** python (>= 2.7), sklern, numpy, pandas, h5py

**git\_url** <https://git.bioconductor.org/packages/BiocSklern>

**git\_branch** RELEASE\_3\_9

**git\_last\_commit** 6682693

**git\_last\_commit\_date** 2019-05-02

**Date/Publication** 2019-10-15

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h5mat	<i>create a file connection to HDF5 matrix</i>
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**Description**

create a file connection to HDF5 matrix

**Usage**

```
h5mat(file, dsname = "assay001")
```

**Arguments**

file	a pathname to an HDF5 file
dsname	internal name of HDF5 matrix to use

**Value**

instance of (S3) `h5py._hl.files.File`

**Examples**

```
fn = system.file("ban_6_17/assays.h5", package="BiocSklern")
h5mat(fn)
```

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H5matref	<i>obtain an HDF5 dataset reference suitable for handling as numpy matrix</i>
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---

**Description**

obtain an HDF5 dataset reference suitable for handling as numpy matrix

**Usage**

```
H5matref(filename, dsname = "assay001")
```

**Arguments**

filename	a pathname to an HDF5 file
dsname	internal name of HDF5 matrix to use, defaults to 'assay001'

**Value**

instance of (S3) `"h5py._hl.dataset.Dataset"`

**Examples**

```

fn = system.file("ban_6_17/assays.h5", package="BiocSklern")
ban = H5matref(fn)
ban
np = import("numpy", convert=FALSE) # ensure
ban$shape
np$take(ban, 0:3, 0L)
fullpca = skPCA(ban)
dim(getTransformed(fullpca))
ta = np$take
# project samples
## Not run: # on celaya2 this code throws errors, and
# I have seen
# .../lib/python2.7/site-packages/sklearn/decomposition/incremental_pca.py:271: RuntimeWarning: Mean of empty
# explained_variance[self.n_components:].mean()
# .../lib/python2.7/site-packages/numpy/core/_methods.py:85: RuntimeWarning: invalid value encountered in dou
# ret = ret.dtype.type(ret / rcount)
ta(ban, 0:20, 0L)$shape
st = skPartialPCA_step(ta(ban, 0:20, 0L))
st = skPartialPCA_step(ta(ban, 21:40, 0L), obj=st)
st = skPartialPCA_step(ta(ban, 41:63, 0L), obj=st)
oo = st$transform(ban)
dim(oo)
cor(oo[,1:4], getTransformed(fullpca)[,1:4])

## End(Not run) # so blocking this part of example for now

```

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SkDecomp-class

*container for sklearn objects and transforms*


---

**Description**

container for sklearn objects and transforms

**Usage**

```

## S4 method for signature 'SkDecomp'
getTransformed(x)

## S4 method for signature 'SkDecomp'
pyobj(x)

```

**Arguments**

x                   instance of SkDecomp

**Value**

the getTransformed method returns a matrix

**Slots**

transform stored as R matrix  
 method string identifying method  
 object reference to the python object with decomposition components

---

skIncrPCA                    *use sklearn IncrementalPCA procedure*

---

**Description**

use sklearn IncrementalPCA procedure

**Usage**

```
skIncrPCA(mat, n_components, batch_size)
```

**Arguments**

mat                    a matrix – can be R matrix or numpy.ndarray  
 n\_components        number of PCA to retrieve  
 batch\_size         number of records to use at each iteration

**Value**

matrix with rotation

**Examples**

```
irloc = system.file("csv/iris.csv", package="BiocSklern")
irismat = SklearnEls()$np$genfromtxt(irloc, delimiter=',')
ski = skIncrPCA(irismat)
ski25 = skIncrPCA(irismat, batch_size=25L) # non-default
getTransformed(ski)[1:3,]
getTransformed(ski25)[1:3,]
```

---

skIncrPPCA                    *optionally fault tolerant incremental partial PCA for projection of samples from SummarizedExperiment*

---

**Description**

optionally fault tolerant incremental partial PCA for projection of samples from SummarizedExperiment

**Usage**

```
skIncrPPCA(se, chunksize, n_components, assayind = 1,
  picklePath = "./skIdump.pkl", matTx = force, ...)
```

**Arguments**

se	instance of SummarizedExperiment
chunksize	integer number of samples per step
n_components	integer number of PCs to compute
assayind	not used, assumed set to 1
picklePath	if non-null, incremental results saved here via <code>sklearn.externals.joblib.dump</code> , for each chunk. If NULL, no saving of incremental results.
matTx	a function defaulting to <code>force()</code> that accepts a matrix and returns a matrix with identical dimensions, e.g., <code>function(x) log(x+1)</code>
...	not used

**Value**

python instance of `sklearn.decomposition.incremental_pca.IncrementalPCA`

**Note**

Will treat samples as records and all features (rows) as attributes, projecting. to an `n_components`-dimensional space. Method will acquire chunk of assay data and transpose before computing PCA contributions. In case of crash, restore from `picklePath` using `SklearnEls()$joblib$load` after loading `reticulate`. You can use the `n_samples_seen_` component of the restored python reference to determine where to restart. You can manage resumption using `skPartialPCA_step`.

**Examples**

```
# demo SE made with TENxGenomics:
# mm = matrixSummarizedExperiment(h5path, 1:27998, 1:750)
# saveHDF5SummarizedExperiment(mm, "tenx_750")
#
if (requireNamespace("HDF5Array")) {
  se750 = HDF5Array::loadHDF5SummarizedExperiment(
    system.file("hdf5/tenx_750", package="BiocSklearn"))
  lit = skIncrPPCA(se750[, 1:50], chunksize=5, n_components=4)
  round(cor(pypc <- lit$transform(dat <- t(as.matrix(assay(se750[,1:50]))))),3)
  rpc = prcomp(dat)
  round(cor(rpc$x[,1:4], pypc), 3)
}
```

---

skKMeans

*interface to `sklearn.cluster.KMeans` with attention to direct work with HDF5*

---

**Description**

interface to `sklearn.cluster.KMeans` with attention to direct work with HDF5

**Usage**

```
skKMeans(mat, ...)
```

**Arguments**

mat                    a matrix-like datum or reference to such  
 ...                    arguments to sklearn.cluster.KMeans

**Note**

You can use `'py_help(SklearnEls())$skcl$KMeans'` to get python documentation on parameters and return structure.

**Examples**

```
# start with numpy array reference as data
irloc = system.file("csv/iris.csv", package="BiocSklearn")
skels = SklearnEls()
irismat = skels$np$genfromtxt(irloc, delimiter=',')
ans = skKMeans(irismat, n_clusters=2L)
names(ans) # names of available result components
table(iris$Species, ans$labels_)
# now use an HDF5 reference
irh5 = system.file("hdf5/irmat.h5", package="BiocSklearn")
fref = skels$h5py$File(irh5)
ds = fref$`__getitem__`("quants") # thanks Samuela Pollack!
ans2 = skKMeans(skels$np$array(ds)$T, n_clusters=2L) # HDF5 matrix is transposed relative to python array layout
table(ans$labels_, ans2$labels_)
ans3 = skKMeans(skels$np$array(ds)$T,
  n_clusters=8L, max_iter=200L,
  algorithm="full", random_state=20L)
```

---

 SklearnEls

*mediate access to python modules from sklearn.decomposition*


---

**Description**

mediate access to python modules from sklearn.decomposition

**Usage**

```
SklearnEls()
```

**Value**

list of (S3) "python.builtin.module"

**Note**

Returns a list with elements np (numpy), pd (pandas), h5py (h5py), skd (sklearn.decomposition), joblib (sklearn.externals.joblib), each referring to python modules.

**Examples**

```
els = SklearnEls()
names(els$skd) # slow at first
# try py_help(els$skd$PCA) # etc.
```

---

skPartialPCA_step	<i>take a step in sklearn IncrementalPCA partial fit procedure</i>
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---

**Description**

take a step in sklearn IncrementalPCA partial fit procedure

**Usage**

```
skPartialPCA_step(mat, n_components, obj)
```

**Arguments**

mat	a matrix – can be R matrix or numpy.ndarray
n_components	number of PCA to retrieve
obj	sklearn.decomposition.IncrementalPCA instance

**Value**

trained IncrementalPCA reference, to which 'transform' method can be applied to obtain projection for any compliant input

**Note**

if obj is missing, the process is initialized with the matrix provided

**Examples**

```
irloc = system.file("csv/iris.csv", package="BiocSklern")
irismat = SklearnEls()$np$genfromtxt(irloc, delimiter=',')
ta = SklearnEls()$np$take
ipc = skPartialPCA_step(ta(irismat,0:49,0L))
ipc = skPartialPCA_step(ta(irismat,50:99,0L), obj=ipc)
ipc = skPartialPCA_step(ta(irismat,100:149,0L), obj=ipc)
head(names(ipc))
ipc$transform(ta(irismat,0:5,0L))
fullproj = ipc$transform(irismat)
fullpc = prcomp(data.matrix(iris[,1:4]))$x
round(cor(fullpc,fullproj),3)
```

---

skPCA	<i>use sklearn PCA procedure</i>
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---

**Description**

use sklearn PCA procedure

**Usage**

```
skPCA(mat, ...)
```

**Arguments**

mat                    a matrix – can be R matrix or numpy.ndarray  
...                    additional parameters passed to sklearn.decomposition.PCA, for additional information use `py_help(SklearnEls())$sk$PCA`

**Value**

matrix with rotation

**Note**

If no additional arguments are passed, all defaults are used.

**Examples**

```
irloc = system.file("csv/iris.csv", package="BiocSklern")
irismat = SklearnEls()$np$genfromtxt(irloc, delimiter=',')
skpi = skPCA(irismat)
getTransformed(skpi)[1:5,]
```



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