Package 'quantsmooth'

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Type Package

Title Quantile smoothing and genomic visualization of array data

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Depends R(>= 2.10.0), quantreg, grid

Description Implements quantile smoothing as introduced in: Quantile smoothing of array CGH data; Eilers PH, de Menezes RX; Bioinformatics. 2005 Apr 1;21(7):1146-53.

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R topics documented:

`chrom.bands` *Dataset of human chromosomes and their banding patterns*

Description

Dataset used to produce human chromosomal ideograms for plotting purposes.

Usage

```
data(chrom.bands)
```

Format

A data frame with 4068 observations on the following 12 variables.

`chr` a character vector

`arm` a character vector

`band` a character vector

`ISCN.top` a numeric vector

`ISCN.bot` a numeric vector

`bases.top` a numeric vector

`bases.bot` a numeric vector

`stain` a character vector

`cM.top` a numeric vector

`cM.bot` a numeric vector

`n.markers` a numeric vector

`p.markers` a numeric vector

Details

The original file gives only the physical map positions. The genetic map positions are interpolated from the Rutgers linkage map (Kong et al 2004).

Source

ftp://ftp.ncbi.nlm.nih.gov/genomes/H_sapiens/maps/mapview/BUILD.35.1/ideogram.gz.

References

Kong X, Murphy K, Raj T, He C, White PS, Matise TC. 2004. A Combined Linkage-Physical Map of the Human Genome. American Journal of Human Genetics, 75(6):1143-8.

Chromosome14 *Example data from several quantitative genomic methods*

Description

A collection of arrays that contains data of chromosome 14 of 3 colorectal tumors. The first tumor shows 1 region of loss, the second tumor shows no abberation, while the third tumor shows loss of 1 copy of the chromosome.

affy.cn Copy number values of 358 probes from Affymetrix 10K genechip. Data was obtained from DChip

affy.pos corresponding probe positions

bac.cn Copy number values of 112 probes from a 1 mb spaced BAC array-CGH

bac.pos corresponding probe positions

ill.cn Copy number values of 207 probes from Illumina GoldenGate Linkage IV data

ill.pos corresponding probe positions

Usage

```
data(chr14)
```

Format

Matrices of copy number values and vectors of chromosomal probe positions

Author(s)

Jan Oosting

drawSimpleChrom *Draw chromosome-like icons*

Description

This function paints chromosomal icons on an existing plot

Usage

```
drawSimpleChrom(x, y, len = 3, width = 1, fill, col, orientation = c("h", "v"), centromere.size = 0.6
```

Arguments

x	start x-position
y	start y-position
len	total length of the chromosome
width	width of the chromosome
fill	character, {"a","p","q","q[1-3]","p[1-3]" }. Events to a chromosome can be depicted by coloring "a"ll of the chromosome, the complete p or q-arm, or a subsegment of the arms
col	color(s) of `fill`
orientation	either "h"orizontal or "v"ertical
centromere.size	The size of the centromere as fraction of the width

Value

This function is executed for its side effects

Author(s)

Jan Oosting

Examples

```
plot(c(0,4),c(0,3),type="n",xaxt="n",yaxt="n",xlab="",ylab="")
drawSimpleChrom(2,3,fill=c("p","q3"),col=c("red","blue"),orientation="v")
```

---

getChangedRegions *getChangedRegions*

Description

retrieve regions of interest in a vector of intensities using quantile smoothing

Usage

```
getChangedRegions(intensities, positions, normalized.to=1, interval, threshold, minlength=2, ...)
```

Arguments

intensities	numeric vector
positions	numeric vector of the same length as intensities. If this argument is not given the results contain the indexes of the `intensities` vector, else the values in `positions` are used. Both vectors are sorted in the order of `positions`.
normalized.to	numeric, reference value. Changes are compared to this value
interval	numeric [0,1], bandwidth around reference. If the smoothed line at the higher quantile drops below the `normalized.to` value, a deleted region is recognized, and vice versa.
threshold	numeric, if the median smoothed value drops below `normalized.to -threshold`, or above `normalized.to + threshold` a changed region is called
minlength	integer, not used currently
...	extra arguments for `quantsmooth` function

Details

This function uses quantsmooth to detect regions in the genome that are abnormal. If interval is set then a smoothed line is calculated for tau = 0.5 -interval/2, and a region is determined as upregulated if this line is above the reference. Down regulation is determined when the smoothed line for tau = 0.5 + interval/2 is below the reference value. If threshold is set then a smoothed line is calculated for tau = 0.5 and up- or down regulation are determined when this line is outside the range [normalized.t -threshold:normalized.to + threshold]

Value

A data.frame with 3 colums is returned. Each row contains a region with columns up, start and end. start and end indicate positions in the vector of the first and last position that were up- or downregulated

Author(s)

Jan Oosting

Examples

```
data(chr14)
getChangedRegions(ill.cn[,1],ill.pos,normalized.to=2,interval=0.5)
```

---

getLambdaMin *getLambdaMin*

Description

Test a set of smoothing parameters to find best fit to data

Usage

```
getLambdaMin(intensities,lambdas,...)
```

Arguments

intensities	numeric vector
lambdas	numeric vector; see quantsmooth
...	extra parameters for quantsmooth.cv; currently only ridge.kappa

Details

Cross validation is performed using a set of lambda values in order to find the lambda value that shows the best fit to the data.

Value

This function returns the lambda value that has the lowest cross validation value on this dataset

Author(s)

Jan Oosting

See Also

quantsmooth.cv

Examples

```
data(chr14)
lambdas<-2^seq(from=-2,to=5,by=0.25)
getLambdaMin(bac.cn[,1],lambdas)
```

---

grid.chromosome *Draw a chromosome using the grid package*

Description

A chromosme is drawn including the cytobands

Usage

```
grid.chromosome(chrom, side = 1, units = "hg19", chrom.width = 0.5, length.out,
        bands = "major", legend = c("chrom", "band", "none"), cex.leg = 0.7, bleach = 0, ...)
```

Arguments

chrom	numeric or character, id of chromosome to plot
side	numeric [1:4], side of rectangle to draw, 4 sides, side 2 and 4 are vertical
units	character or data.frame, type of units for genomic data, or a dataframe with UCSC cytoband data, see lengthChromosome
chrom.width	numeric [0,1], The width relative to the width (sides 2 and 4) or height(sides 1 and 3) of the viewport
length.out	numeric, size of native units of viewport
bands	character, draw either major or minor bands
legend	character, type of legend
cex.leg	numeric, relative size of legend text
bleach	numeric [0,1], proportion by which to bleach the chromosome
...	arguments for viewport(), especially x,y, width, and height

Details

The chromosome is drawn within a rectangle defined by x, y, width, and height, which is pushed as a viewport. The legend is drawn within the same rectangle in the space left over by chrom.width.

Value

This function is executed for its side effects

Author(s)

David L Duffy ,Jan Oosting

References

lodplot package

See Also

paintCytobands

Examples

```
grid.newpage()
grid.chromosome(1,units="bases",height=0.15)
```

`lengthChromosome` *Retrieve chromosomal length*

Description

Retrieve human chromosomal length from NCBI data

Usage

```
lengthChromosome(chrom, units = "hg19")
```

Arguments

`chrom`	vector of chromosomal id, 1:22,X,Y
`units`	character, or data.frame, see details

Details

The cytoband data was originally obtained from the lodplot package by David Duffy, which contained basepair data from genome version hg17, but also the linkage related positions in cM. These datasets have units `"bases"` and `"cM"` respectively. Cytoband data for genome versions "hg18", "hg19", "hg38" and "mm10" has been included, and can be referenced by these strings. It is also possible to use cytoband data as obtained from the UCSC site, by downloading the `cytoBand.txt.gz` or `cytoBandIdeo.txt.gz` annotation file for a species (see example below). Note however that this information is not available for most species.

Value

A numeric vector in the requested units

Author(s)

Jan Oosting

Examples

```
# Show length of chromosome 1 in several types of units
lengthChromosome(1,"cM")
lengthChromosome(1,"bases")
lengthChromosome(1,"hg38")
# mm9 cytoband data
temp <- tempfile(fileext = ".txt.gz")
download.file("http://hgdownload.soe.ucsc.edu/goldenPath/mm9/database/cytoBand.txt.gz",temp)
mm9cytobands <- read.table(temp,sep="\t")
lengthChromosome(1,mm9cytobands)
# remove temp file
unlink(temp)
```

---

numericCHR *Conversion of chromosome IDs between numeric and character*

Description

The function converts chromosomal ids to their numeric form, and the sex chromosomes to values between 98 and 100. This simplifies sorting on chromosome ID

Usage

```
numericCHR(CHR, prefix="chr")
characterCHR(CHR, prefix="")
```

Arguments

CHR	character/numeric vector for both functions the mode of the input is not forced. For numericCHR strings "X","Y" and "XY" are converted to 98,99 and 100 respectively.
prefix	character, string is excluded from (numericCHR) or prepended to (characterCHR) all items of the output

Value

numericCHR returns a numeric vector of same length as CHR characterCHR returns a character vector of same length as CHR

Author(s)

Jan Oosting

Examples

```
chroms<-c("3","2","8","X","7","Y","5","1","9","10","11","12","4","6")
sort(chroms)
sort(numericCHR(chroms))
characterCHR(sort(numericCHR(chroms)),prefix="chr")
```

paintCytobands *Paint a chromosomal idiogram*

Description

Paints a human chromosomal idiogram in an existing plot Adapted from the paint.chromosome function in the lodplot package by David L Duffy

Usage

```
paintCytobands(chrom, pos = c(0, 0), units = "hg19", width = 0.4,
           length.out, bands = "major", orientation = c("h","v"), legend = TRUE,
            cex.leg = 0.7, bleach = 0, ...)
```

Arguments

chrom	chromosomal id, chromosome to plot 1:22,X,Y
pos	numeric vector of length 2, position in the plot to start the plot
units	character or data.frame, type of units for genomic data, or a dataframe with UCSC cytoband data, see lengthChromosome
width	numeric, width of the chromosome, the chromosome is plotted between pos[2] and pos[2]-width
length.out	numeric, if given, the chromosome will have this length in the plot
bands	if not equal to "major", then also the minor bands will be plotted
orientation	chromosome is plotted either *H*orizontally to the right of the starting point or *V*ertically down from the starting point
legend	logical, if TRUE then the bandnames are plotted next to the chromosome
cex.leg	numeric, relative size of legend text
bleach	numeric [0,1], proportion by which to bleach the chromosome
...	extra parameters for plot

Value

This function is executed for its side effects

Author(s)

David L Duffy , Jan Oosting

References

lodplot package

Examples

```
plot(c(0,lengthChromosome(14,"bases")),c(-2,2),type="n",xaxt="n",yaxt="n",xlab="",ylab="")
 paintCytobands(14,units="bases")
```

plotChromosome *Wrapper for plotSmoothed*

Description

This function is a wrapper for plotSmoothed, to make data subsetting easier

Usage

```
plotChromosome(gendata, chrompos, chromosome, dataselection = NULL, ylim = NULL, normalized.to = NUL
```

Arguments

gendata	numeric matrix or data.frame
chrompos	chrompos object with same numer of rows as gendata
chromosome	numeric, chromosme to show
dataselection	optional, subset of samples/columns in gendata
ylim	limits for plot
normalized.to	y-value(s) for line
grid	x-value(s) for line
smooth.lambda	smoothing parameter, see quantsmooth
interval	position of extra lines besides median, see plotSmoothed
...	extra arguments for plotSmoothed

Value

The function is used for its side effects

Author(s)

Jan Oosting

See Also

plotSmoothed, quantsmooth

plotSmoothed *plotSmoothed*

Description

Plot a smoothed line together with the original data values

Usage

```
plotSmoothed(intensities, position, ylim=NULL, ylab="intensity", xlab="position", normalized.to=
```

Arguments

intensities	numeric vector or matrix, data are plotted by column
position	numeric vector; the length should be the number of rows in intensities
ylim	numeric vector of length 2, limits for plot. If NULL then the minimal and maximal value in intensities is used
ylab	character, label for y-position
xlab	character, label for x-position
normalized.to	numeric, a line(s) is drawn at this horizontal position
grid	numeric, a line(s) is drawn at this vertical position
smooth.lambda	numeric, smoothing parameter see quantsmooth
interval	numeric (0..1), plotting of extra smoothed lines around median. With interval = 0.5 the 0.25 and 0.75 quartiles are plotted, with interval = 0.9 the 0.05 and 0.95 quantiles are plotted,
plotnew	logical, if TRUE a new plot is created, else the data are plotted into an existing plot
cols	color vector, colors for columns in intensities
cex.pts	size of the dots in the plot. Set to 0 to skip plotting the dots
...	extra parameters for plot

Details

This function plots the raw data values as dots and the median smoothed values as a continuous line. If interval is supplied these are plotted as lines in different line types. More than 1 interval can be given.

Value

This function is used for its side effects

Author(s)

Jan Oosting

See Also

quantsmooth

Examples

```
data(chr14)
plotSmoothed(bac.cn,bac.pos,ylim=c(1,2.5),normalized.to=2,smooth.lambda=2.5)
```

`position2Cytoband` *Determine cytoband position based on location of probe*

Description

Determine cytoband position based on location of probe

Usage

```
position2Cytoband(chrom, position, units = "hg19", bands = c("major", "minor"))
```

Arguments

`chrom`	chromosomal id, chromosome to plot 1:22,X,Y
`position`	numeric vector
`units`	character, type of positional unit
`bands`	chararcter, type of cytoband

Value

Character vector with cytobands, if an illegal position was used, the value "-" is returned. All positions within a single function call should be for a single chromosome

Author(s)

Jan Oosting

See Also

`lengthChromosome`

Examples

```
position2Cytoband(1,c(50e6,125e6,200e6),units="bases")
position2Cytoband(1,c(50,125,200),units="cM",bands="minor")
```

prepareGenomePlot *Set up a full genome plot*

Description

This function starts up a plot consisting of all chromosomes of a genomen, including axes with chromosome names.

Usage

```
prepareGenomePlot(chrompos, cols = "grey50", paintCytobands = FALSE, bleach = 0, topspace = 1, organ
                  sexChromosomes = FALSE, units = "hg19",...)
```

Arguments

chrompos	chrompos object, data.frame with CHR column identifying the chromosome of probes, and a MapInfo column identifying the position on the chromosome
cols	color(s) for the chromosome lines
paintCytobands	logical, use paintCytoband to plot ideograms for all chromosomes
bleach	numeric [0,1], proportion by which to bleach the ideograms
topspace	numerical, extra space on top of plot, i.e. for legends
organism	character, if given a 2 column plot is created with the chromosomes for the given species. Currently "hsa", "mmu", and "rno" are supported
sexChromosomes	logical, if TRUE then also the sex chromosomes X and Y are plotted
units	characterr or data.frame, type of units for genomic data, or a dataframe with UCSC cytoband data, see lengthChromosome
...	extra arguments for plot function

Details

If organism is not supplied then a single column is plotted of the available chromosomes in chrompos$CHR. The arguments paintCytobands, bleach, and sexChromosomes are not used in that case. If organism is supplied and chrompos is NULL then a result is generated with the starting Y and X position of each chromosome

Value

A matrix with 2 columns that contain the Y and X positions for the probes on the plot

Author(s)

Jan Oosting

quantsmooth *quantsmooth*

Description

Quantile smoothing of array data

Usage

```
quantsmooth(intensities,smooth.lambda=2, tau=0.5, ridge.kappa=0,smooth.na=TRUE,segment)
```

Arguments

intensities	numeric vector
smooth.lambda	numeric
tau	numeric [0..1], the quantile desired; see rq.fit
ridge.kappa	fudge parameter; see details
smooth.na	logical; handling of NA
segment	integer, length of overlapping segments

Value

This function returns a vector of the same length as intensities, or a matrix if the length of tau is greater than 1.

Author(s)

Jan Oosting

Examples

```
data(chr14)
plot(quantsmooth(bac.cn[,1],smooth.lambda=2.8),type="l")
```

---

quantsmooth.cv *quantsmooth.cv*

Description

Cross validation of smoothing parameters

Usage

```
quantsmooth.cv(intensities,smooth.lambda=2, ridge.kappa=0)
```

Arguments

intensities	numeric vector
smooth.lambda	numeric; see quantsmooth
ridge.kappa	fudge parameter; see quantsmooth

Details

Cross validation is performed by calculating the fit from the even indices on the odd indices and vice versa.

Value

This function returns the sum of squared differences or NA if the fitting function gave an error

Author(s)

Jan Oosting

See Also

getLambdaMin

Examples

```
data(chr14)
# A low value is indicative of a better fit to the data
quantsmooth.cv(bac.cn[,1],1)
quantsmooth.cv(bac.cn[,1],2.8)
```

---

quantsmooth.seg *quantsmooth.seg*

Description

segmented Quantile smoothing of array data

Usage

```
quantsmooth.seg(y, x = 1:length(y), lambda = 2, tau = 0.5,kappa = 0, nb = length(x))
```

Arguments

y	numeric vector
x	numeric vector of same length as y. Position of values
lambda	numeric
tau	numeric [0..1], the quantile desired; see rq.fit
kappa	fudge parameter; see details
nb	integer, basis

Value

This function returns a vector of the same length as y

Author(s)

Jan Oosting

Examples

```
data(chr14)
plot(quantsmooth.seg(bac.cn[,1],lambda=2.8,nb=50),type="l")
```

`scaleto` *Scales data within a range to a new range*

Description

This function scales data to a new range while enforcing the boundaries. This can be helpful in preventing overlap between chromosomal plots that display multiple chromosomes in the same plot

Usage

```
scaleto(x, fromlimits = c(0, 50), tolimits = c(0.5, -0.5), adjust = TRUE)
```

Arguments

`x`	numeric
`fromlimits`	numeric vector with length 2, original range of data
`tolimits`	numeric vector with length 2, target range of data
`adjust`	logical, if `TRUE` then the target values are clipped to the target range

Value

numeric of same size as `x`

Author(s)

Jan Oosting

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